

Antimicrobial Susceptibility Patterns of Carbapenem-Resistant *Klebsiella pneumoniae* Causing Lower Respiratory Infections

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ABSTRACT

Introduction: Lower respiratory tract infections (LRTIs) continue to be a significant health burden, and the most common bacterial causes include multi-drug resistant (MDR) bacteria like *Klebsiella pneumoniae* (*K. pneumoniae*) infections. The emergence of carbapenem-resistant *K. pneumoniae* (CRKP) has added to these concerns due to limited treatment options and poorer clinical outcomes. This study was aimed to find out antimicrobial susceptibility pattern and to detect mechanism of carbapenem resistance among *K. pneumoniae* isolates from LRTI patients. **Methods & Materials:** This cross-sectional study was performed to find out the antimicrobial susceptibility pattern of carbapenem resistant *Klebsiella pneumoniae* (CRKP) causing lower respiratory tract infection (LRTIs). This study was performed in the Department of Microbiology of Sylhet MAG Osmani Medical College, Sylhet, Bangladesh from July, 2021 to June, 2022. A total of 153 sputum samples were received from the diagnosed cases of LRTIs from adult patients (aged ≥ 18 years). Data analysis performed by Statistical Package for Social Sciences (SPSS) version 26. **Result:** A total of 153 sputum samples were analyzed, with bacterial growth detected in 66.0% of cases. *Klebsiella pneumoniae* was isolated in 20.9% of samples, of which 25.0% were carbapenem-resistant. CRKP isolates showed high rates of multi-drug resistance, particularly against amoxicillin-clavulanic acid, ciprofloxacin, ceftazidime, azithromycin, meropenem, imipenem, and cefepime. Carbapenemase production was detected in 75.0% of CRKP isolates by Modified Hodge Test. **Conclusion:** This research revealed the extent of the problem of carbapenem-resistant *Klebsiella pneumoniae* among

LRTI cases and a substantial number of isolates found to be carbapenemase producers. The isolates were multi-drug resistant to commonly used antibiotics like β -lactams, fluoroquinolones, and most of the other routinely administered drugs, and therapeutic options were almost nil.

Keywords: Antimicrobial Susceptibility, Carbapenem Resistant, *Klebsiella Pneumoniae*, LRTI

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INTRODUCTION

Lower respiratory tract infections (LRTIs) are a common and significant cause of morbidity and mortality worldwide. They are also responsible for lots of hospital admissions, mainly amongst critically ill and immunocompromised patients. Bacterial causes of LRTI are common, and *Klebsiella pneumoniae* has become an increasingly significant opportunistic pathogen in the community, in hospitals, causing pneumonia, ventilator-associated pneumonia, and fulminant respiratory infections [1,2]. *Klebsiella pneumoniae* is a Gram-negative bacillus of the Enterobacteriaceae family. Its virulence lies in its polysaccharide capsule that helps in evading the host immune system [3]. Over the past decade, the appearance of an antimicrobial resistance pattern among the etiological agent *Klebsiella pneumoniae* has become one of the most challenging issues for global health community. Of the multiple forms of resistance developed by the organism, the production of carbapenemase enzymes and the resultant

resistance to carbapenems have been raised to a much higher level than other mechanisms of antibiotic inactivation because of carbapenems are considered as drugs of choice for treatment of severe infections caused by multidrug resistant (MDR) gram-negative bacteria [4]. During this period the emergence of carbapenem-resistant *K. pneumoniae* (CRKP) has led to a dramatic decline in therapeutic options and high rates of clinical failures, prolonged hospital stays, increased healthcare costs, and high mortality [5]. The mortality rate in patients infected with CRKP has been reported to be greater than 40% which is much higher than with infections due to carbapenem sensitive strains [6]. CRKP infections have been related to high morbidity and mortality in groups of patients in the intensive care unit as well as those admitted in hospitals with APACHE II scores and having invasive procedures and prior use of antibiotics [7]. Resistance conferred by CRKP can be achieved by a number of methods including production of a range of carbapenemase enzymes like

Klebsiella pneumoniae carbapenemase (KPC), new Delhi B-lactamase (NDM), VIM (Verona integron-encoded Metallo-lactamase), OXA-48 and also by loss of porins and increased efflux [8]. These resistance determinants are frequently plasmid-mediated, facilitating rapid dissemination among bacterial populations and contributing to hospital outbreaks [9]. Global surveillance studies have demonstrated an increasing prevalence of CRKP in both developed and developing countries, emphasizing the urgent need for continuous monitoring and antimicrobial stewardship interventions [10]. Recent meta-analytic evidence estimated that approximately 28.7% of hospital-acquired *K. pneumoniae* infections globally are carbapenem-resistant. Antimicrobial susceptibility profiles may vary based on geographic location, healthcare setting and resistant genes being circulated. As a result, institution specific surveillance studies are really important in clinical management and infection control policies. Assessment of susceptibility pattern of carbapenem

resistant *Klebsiella pneumoniae* from LRTIs shall give an idea about prevailing resistance trends and help to conserve antimicrobial resources. To that extent, present study was undertaken to report antimicrobial susceptibility pattern of carbapenem resistant *Klebsiella pneumoniae* from LRTIs.

METHODS & MATERIALS

This is a cross-sectional study, which was carried out in the Department of Microbiology, Sylhet MAG Osmani Medical College Sylhet Bangladesh, during the period of July, 2021 to June, 2022 to determine the pattern of antimicrobial susceptibility in carbapenem resistant *Klebsiella pneumoniae*, isolated from LRTIs. One hundred and fifty-three sputum samples were collected from adult patients (≥ 18 years of age), with LRTIs attending the Medicine outpatient department (OPD) and the clinical wards of Sylhet MAG Osmani Medical College Hospital, Sylhet. Immunosuppressive therapy users and cases

of tuberculosis were excluded. Sputum specimens were collected in sterile, clean, leak-proof, wide-necked containers and transported promptly to the microbiology laboratory for immediate processing. Macroscopic and microscopic examinations of the specimens were performed before culture on Blood agar and MacConkey agar media, followed by incubation at 37°C for 24 hours. Suspected *Klebsiella pneumoniae* isolates were identified through gram staining technique and standard biochemical tests, including indole, catalase, urease, citrate, oxidase, and triple sugar iron (TSI) tests. Antimicrobial susceptibility testing (AST) of the isolates was carried out by the Kirby Bauer Disc Diffusion method on Mueller Hinton agar media, following the Clinical and Laboratory Standards Institute (CLSI) guideline, 2021. Phenotypic detection of carbapenemase production was carried out by the Modified Hodge Test (MHT). A 0.5 McFarland suspension of *E. coli* ATCC 25922 was prepared and swarm inoculum

was evenly spread using a sterile swab on the surface of Muller Hinton agar media and a disk of meropenem (10 g) was placed in the center of the plate and the test isolate streaked radially from the disk border to the edge of the plate. Plates were incubated at 37°C for 24 hours and interpreted as per CLSI 2021 guidelines. Prior Ethical approval was obtained from Ethical Review Committee Sylhet MAG Osmani Medical College for this study. All data were entered and analyzed using Statistical Package for Social Sciences (SPSS) version 26. Only descriptive statistical analyses were performed.

RESULTS

Among the 153 sputum samples collected in this study, the participants had a mean age of 52.43 ± 17.12 years, ranging from 18 to 87 years. Male participants constituted the majority (69.3%), with a male-to-female ratio of 2.3:1. (Table I).

Table I

Demographic Characteristics and Microbiological Findings of Study Participants ($n=153$).

Variables	Frequency (n)	Percentage (%)
Age (years)		
Mean \pm SD	52.43 \pm 17.12	—
Range	18–87	—
Gender		
Male	106	69.3
Female	47	30.7
Male: Female ratio	2.3: 1	—

A total of 153 sputum samples were collected from the study participants. Among them, 101 (66.0%) samples showed positive bacterial growth on culture, while

52 (34.0%) samples demonstrated no microbial growth. Of the total samples, single bacterial growth was identified in 87 (56.8%) cases, whereas mixed bacterial

growth was observed in 14 (9.2%) cases (Table II).

Table II

Sputum Culture Findings Among the Study Participants ($n = 153$).

Sputum Culture Findings	Frequency (n)	Percentage (%)
Total sputum samples collected	153	100.0
Growth positive	101	66.0
No growth	52	34.0
Single bacterial growth	87	56.8
Mixed bacterial growth	14	9.2

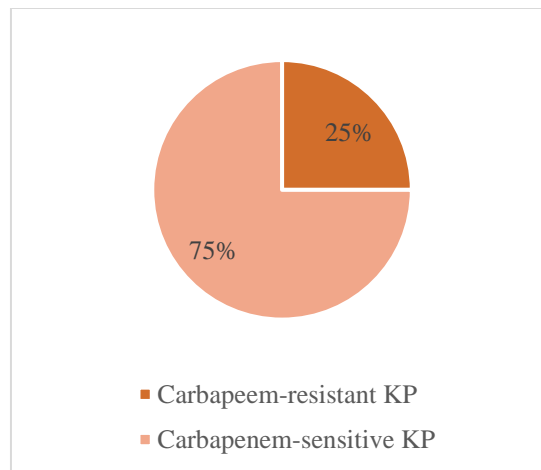


Figure 1 Pie chart showing positive *Klebsiella pneumoniae* (KP) isolates (n=32).

Among the isolated organisms, *Klebsiella pneumoniae* was detected in 32 (20.9%) cases. Of these isolates, 8 (25.0%) were identified as carbapenem-resistant *Klebsiella pneumoniae* (CRKP), whereas 24 (75.0%) were carbapenem-sensitive strains (Figure 1).

The highest sensitivity was observed for piperacillin–tazobactam, ceftazidime,

cefotaxime, and amikacin, each showing sensitivity in 4 (50.0%) isolates. Moderate sensitivity was found for ceftaxitin, meropenem, imipenem, azithromycin, and cefepime, while the lowest sensitivity was observed for amoxicillin–clavulanic acid and ciprofloxacin, with only 1 (12.5%) sensitive isolate each. High resistance rates were noted against amoxicillin–clavulanic acid and ciprofloxacin (87.5% each),

followed by ceftaxitin, meropenem, imipenem, and azithromycin (75.0% each). Intermediate susceptibility was identified only for piperacillin–tazobactam and amikacin in 1 (12.5%) isolate each, and for cefepime in 2 (25.0%) isolates (Table III).

Table III
Antimicrobial Susceptibility Patterns of Carbapenem-Resistant *Klebsiella pneumoniae* in LRTI (n=8).

Antimicrobials	Sensitive n (%)	Intermediate n (%)	Resistant n (%)
Piperacillin–tazobactam	4 (50.0)	1 (12.5)	3 (37.5)
Ceftazidime	4 (50.0)	0 (0.0)	4 (50.0)
Cefotaxime	4 (50.0)	0 (0.0)	4 (50.0)
Ceftaxitin	2 (25.0)	0 (0.0)	6 (75.0)
Amoxicillin–clavulanic acid	1 (12.5)	0 (0.0)	7 (87.5)
Meropenem	2 (25.0)	0 (0.0)	6 (75.0)
Imipenem	2 (25.0)	0 (0.0)	6 (75.0)
Amikacin	4 (50.0)	1 (12.5)	3 (37.5)
Azithromycin	2 (25.0)	0 (0.0)	6 (75.0)
Cefepime	2 (25.0)	2 (25.0)	4 (50.0)
Ciprofloxacin	1 (12.5)	0 (0.0)	7 (87.5)

DISCUSSION

In this study, among the 32 isolated *Klebsiella pneumoniae* strains, 8 (25.0%) were identified as carbapenem-resistant *Klebsiella pneumoniae* (CRKP), which was comparable to findings reported by Shanmugam et al, Bina et al, and Aminul et al [11,12]. Among the 8 CRKP isolates, carbapenemase production detected by the Modified Hodge Test (MHT) was observed in 75.0% of cases, which was consistent with the findings of Shanmugam et al, where MHT positivity was reported in 86.6% of CRKP isolates [11]. Similar high positivity rates ranging from 71% to 84% have also been reported in other studies [13-15]. The increasing emergence of carbapenemase-producing *K. pneumoniae* in hospital settings may be attributed to several factors, including irrational and excessive antibiotic use, inappropriate dosing practices, circulation of counterfeit or substandard antibiotics, and inadequate

infection control measures related to the isolation and management of multidrug-resistant pathogens. Additionally, these organisms may also be acquired from community sources, as they have been isolated from common water reservoirs [16]. Regarding antimicrobial susceptibility patterns, CRKP isolates in the present study demonstrated high resistance rates to amoxicillin–clavulanic acid (87.5%), ciprofloxacin (87.5%), ceftaxitin (75.0%), azithromycin (75.0%), meropenem (75.0%), imipenem (75.0%), and cefepime (50.0%). Similar multidrug resistance patterns were reported by Bahrami et al, where CRKP isolates exhibited resistance to ciprofloxacin (96.7%), cefepime (100.0%), piperacillin–tazobactam (98.4%), ceftazidime (98.8%), and amikacin (87.0%) [17-18]. Furthermore, carbapenemase-producing *K. pneumoniae* detected by conventional PCR in the present study exhibited extensive resistance to multiple

antimicrobial agents, including colistin (100.0%), amoxicillin–clavulanic acid (100.0%), ceftaxitin (100.0%), ciprofloxacin (100.0%), cefotaxime (75.0%), ceftazidime (75.0%), cefepime (75.0%), azithromycin (75.0%), meropenem (75.0%), and imipenem (75.0%). Resistance among carbapenemase-producing *K. pneumoniae* frequently extends beyond β-lactams to include fluoroquinolones, aminoglycosides, and tetracyclines, creating substantial therapeutic challenges. Although optimal treatment strategies remain uncertain, some evidence suggests that appropriately administered carbapenem-based regimens may still retain clinical value in selected cases [19]. Therefore, routine implementation of the Modified Hodge Test in microbiology laboratories is important, as positive findings can provide clinicians with critical information regarding carbapenemase production. This approach offers a rapid and cost-effective method for

detecting carbapenemase production in *K. pneumoniae*.

LIMITATIONS

- The study was conducted at a single tertiary care centre with a relatively small sample size, limiting the generalizability of the findings.
- Only descriptive statistical analysis was performed, which restricted the assessment of associations between variables.
- Molecular analysis should be done to detect carbapenem resistance genes among the isolates.

CONCLUSION

This study demonstrated a considerable burden of carbapenem-resistant *Klebsiella pneumoniae* among lower respiratory tract infection cases, with a notable proportion of isolates producing carbapenemase enzymes. The isolates showed high levels of multidrug resistance, particularly against commonly used antibiotics such as β -lactams, fluoroquinolones, and other routinely prescribed agents, leaving very limited therapeutic options.

RECOMMENDATION

All clinically significant isolates of *K. pneumoniae* should be subjected to routine antimicrobial susceptibility testing before administering antibiotics. Rapid identification of carbapenemase producing organisms using phenotypic and molecular methods should be strengthened in diagnostic laboratories. Strict compliance to measures for infection prevention and control and implementation of good antimicrobial stewardship programs are the necessary steps to prevent dissemination of multidrug resistant organisms.

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CONFLICT OF INTEREST

None declared.

REFERENCES

1. Miron M, Blaj M, Ristescu AI, Iosep G, Avădanei AN, Iosep DG, Crişan-Dabija R, Ciocan A, Perţea M, Manciu CD, Luca Ş. Hospital-acquired pneumonia and ventilator-associated pneumonia: a literature review. *Microorganisms*. 2024 Jan 20;12(1):213.
2. Magill SS, Edwards JR, Bamberg W, Beldavs ZG, Dumyati G, Kainer MA, Lynfield R, Maloney M, McAllister-Hollod L, Nadle J, Ray SM. Multistate point-prevalence survey of health care-associated infections. *New England Journal of Medicine*. 2014 Mar 27;370(13):1198-208.
3. Paczosa MK, Meecs J. *Klebsiella pneumoniae*: going on the offense with a strong defense. *Microbiology and molecular biology reviews*. 2016 Sep;80(3):629-61.
4. Karampatakis T, Tsergouli K, Behzadi P. Carbapenem-resistant *Klebsiella pneumoniae*: virulence factors, molecular epidemiology, and latest updates in treatment options. *Antibiotics*. 2023 Jan 21;12(2):234.
5. van Loon K, Voor in 't holt AF, Vos MC. A systematic review and meta-analysis of the clinical epidemiology of carbapenem-resistant Enterobacteriaceae. *Antimicrobial agents and chemotherapy*. 2018 Jan;62(1):10-128.
6. Xu L, Sun X, Ma X. Systematic review and meta-analysis of mortality of patients infected with carbapenem-resistant *Klebsiella pneumoniae*. *Annals of clinical microbiology and antimicrobials*. 2017 Mar 29;16(1):18.
7. Martin A, Fahrbach K, Zhao Q, Lodise T. Association between carbapenem resistance and mortality among adult, hospitalized patients with serious infections due to Enterobacteriaceae: results of a systematic literature review and meta-analysis. In *Open Forum Infectious Diseases* 2018 Jul (Vol. 5, No. 7, p. ofy150). US: Oxford University Press.
8. Hansen GT. Continuous evolution: perspective on the epidemiology of carbapenemase resistance among Enterobacterales and other Gram-negative bacteria. *Infectious diseases and therapy*. 2021 Mar;10(1):75-92.
9. Logan LK, Weinstein RA. The epidemiology of carbapenem-resistant Enterobacteriaceae: the impact and evolution of a global menace. *The Journal of Infectious Diseases*. 2017 Feb 15;215(suppl_1):S28-36.
10. Lin XC, Li CL, Zhang SY, Yang XF, Jiang M. The global and regional prevalence of hospital-acquired carbapenem-resistant *Klebsiella pneumoniae* infection: a systematic review and meta-analysis. In *Open Forum Infectious Diseases* 2024 Feb (Vol. 11, No. 2, p. ofad649). US: Oxford University Press.
11. Shanmugam P, Meenakshisundaram J, Jayaraman P. blaKPC gene detection in clinical isolates of carbapenem-resistant Enterobacteriaceae in a tertiary care hospital. *Journal of clinical and diagnostic research: JCDR*. 2013 Dec 15;7(12):2736.
12. Aminul P, Anwar S, Molla MM, Miah MR. Evaluation of antibiotic resistance patterns in clinical isolates of *Klebsiella pneumoniae* in Bangladesh. *Biosafety and Health*. 2021 Dec 30;3(06):301-6.
13. Bina M, Pournajaf A, Mirkalantari S, Talebi M, Irajian G. Detection of the *Klebsiella pneumoniae* carbapenemase (KPC) in *K. pneumoniae* Isolated from clinical samples by the Phenotypic and Genotypic Methods. *Iranian journal of pathology*. 2015;10(3):199.
14. Cury AP, Andreazzi D, Maffucci M, Caiaffa-Junior HH, Rossi F. The modified Hodge test is a useful tool for ruling out *Klebsiella pneumoniae* carbapenemase. *Clinics*. 2012 Dec;67(12):1427-31.
15. Mosca A, Miragliotta L, Del Prete R, Tzakis G, Dalfino L, Bruno F, Pagani L, Migliavacca R, Piazza A, Miragliotta G. Rapid and sensitive detection of bla KPC gene in clinical isolates of *Klebsiella pneumoniae* by a molecular real-time assay. *SpringerPlus*. 2013 Jan 30;2(1):31.
16. Bhaskar BH, Mulki SS, Joshi S, Adhikary R, Venkatesh BM. Molecular characterization of extended spectrum β -lactamase and carbapenemase producing *Klebsiella pneumoniae* from a tertiary care hospital. *Indian journal of critical care medicine: peer-reviewed, official publication of the Indian Society of Critical Care Medicine*. 2019 Feb;23(2):61.
17. Raghunathan A, Samuel L, Tibbetts RJ. Evaluation of a real-time PCR assay for the detection of the *Klebsiella pneumoniae* carbapenemase genes in microbiological samples in comparison with the modified Hodge test. *American journal of clinical pathology*. 2011 Apr 1;135(4):566-71.
18. Bahrami S, Shafiee F, Hakamifard A, Fazeli H, Soltani R. Antimicrobial susceptibility pattern of carbapenemase-producing Gram-negative nosocomial bacteria at Al Zahra hospital, Isfahan, Iran. *Iranian journal of microbiology*. 2021 Feb;13(1):50.
19. Robilotti E, Deresinski S. Carbapenemase-producing *Klebsiella pneumoniae*. *F1000prime reports*. 2014 Sep 4;6:80.