

Relation of Apoptosis Marker CK-18 fragment M30 with hepatic Necroinflammation in patients with HBV related Chronic Liver Disease

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ABSTRACT:

Background and Objective: Caspase-cleaved cytokeratin 18 (CK18 M30) is a potential clinically useful biomarker in liver disease as it is released from hepatocytes during apoptosis. Chronic hepatitis B virus (HBV) infection affects more than 400 million people worldwide. For treatment of chronic hepatitis B, it is very essential to know the necro inflammatory status of liver. Cytokeratin (CK) 18 is an intermediary filament protein, expressed in hepatocytes, which is proteolytically cleaved during liver damage. In this study, we aimed to investigate whether serum CK-18 fragment M30 level significantly related with the hepatic necroinflammatory activity in patients with HBV related compensated chronic liver disease (CLD). **Methods:** This was a prospective observational study. All patients who met the inclusion and exclusion criteria were assessed for liver biopsy. The total sample was 40 patients. This study was conducted in Department of Hepatology, Bangabandhu Sheikh Mujib Medical University. Per cutaneous transthoracic liver biopsy was done. Specimens were sent to department of pathology, BSMMU for METAVIR scoring. **Result:** Among 40 CHB patients, the highest frequency was found at 21-30 age groups, male 31(77.5%) and female 9(22.5%), the mean HBV DNA PCR was found 5.3 ± 1.7 (IU/ml). The mean AST and ALT were found 40.2 ± 20.2 (U/L) and 66.4 ± 68.2 (U/L) respectively. 16 patients were HBeAg positive and 24 patients were HBeAg negative. CK-18 M30 level in both HBeAg negative CHB patients and HBeAg positive patients were almost similar (128.8 ± 32.91 and 123.9 ± 28.1), Activity METAVIR score of hepatic necro inflammation reveals, A0 was 0 (0.0%), A1 was 7(17.5%), A2 was 21(52.5%), A3 was 12(30.0%), Correlation between factors and activity METAVIR score on spearman correlation test reveals, Correlation co-efficient (r value)-age was 0.204, GGT was 0.287, ALT was 0.333, AST was 0.360, Serum CK-

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18 fragment M30 level was negatively correlated with activity score of hepatic necro inflammation ($r = -0.073$; $p = 0.357$). The correlation between ALT and AST with activity score were statistically significant ($p < 0.05$). The area under the receiver-operator characteristic (ROC) curves for prediction of serum CK-18 fragment M30 level reveals - area under curve (AUC) 0.307, which gave a cut off value of 100 U/L with 78.8% sensitivity and 14.3% specificity for prediction of significant necro inflammation (A2 and A3). **Conclusion:** This study indicates there was no correlation between serum CK-18 fragment M30 level and hepatic necroinflammatory activity in patients with HBV related compensated chronic liver disease (CLD). As a result, CK 18 M30 cannot be used as an accurate non-invasive predictor of significant inflammatory activities in patients with CHB.

Key words: Chronic Hepatitis B, Chronic Liver Disease, Necro-inflammatory status, Cytokeratin (CK)-18 fragment M30, Hepatic Fibrosis

(The Planet 2020; 4(1):4-11)

INTRODUCTION:

There are an estimated two billion people with serological markers of present or past Hepatitis B virus (HBV) infection globally; 257 million of these are chronically infected.¹ The outcomes of acute HBV infection range from complete recovery to fulminant liver disease. A failure to clear HBV after acute infection may lead to either inactive or active chronic infection, which can induce hepatic insufficiency, end-stage liver disease including liver cirrhosis (LC) and hepatocellular carcinoma (HCC).^{2,3} Bangladesh is within the intermediate zone of prevalence (5.4%) of HBV infection and HBeAg negative variant is predominant cause of chronic hepatitis in incidentally detected HBsAg positive patient.⁶ Transmission of HBV occurs either perinatally or horizontally. Chronic hepatitis affects the younger and middle aged population of Bangladesh and may lead to development of cirrhosis and hepatocellular carcinoma. This favours horizontal transmission in early childhood contrary to vertical transmission.^{7,8} Chronic hepatitis B diagnosis is based on clinical examination

and laboratory tests. The assessment of the severity of the liver disease includes LFT, HBV DNA, hepatic ultrasound and liver Biopsy.⁵

Patients with significant hepatic inflammation and fibrosis are at the highest risk of complications like cirrhosis of liver and hepatocellular carcinoma.⁹ Histological examination of liver biopsy is the current gold standard for the detection of liver damage. This procedure provides important information regarding the severity of necroinflammatory activity and fibrosis, features potentially useful for predicting treatment response and prognosis.⁹ Liver biopsy has a number of limitations. Liver biopsy is invasive, costly, and limited by sampling error and poor intra and inter observer variability.¹⁰⁻¹² Many recent studies clearly indicate that liver biopsy is prone to sampling errors and may underestimate the extent of liver injury.¹³ Considering these limitations and patient reluctance to undergo liver biopsy, noninvasive predictors of histological severity are desperately needed.

Cytokeratin is the major intermediate filament protein in cells. These are subdivided into type I (acidic) and type II (basic) keratins. Cytokeratin-18 is type I keratin. Keratin expression is tissue specific with different pairs. Adult hepatocytes express keratin 18 (K18; type I) and keratin 8 (K8; type II) exclusively. It forms cytoplasmic network. CK-18 maintains normal cellular and mitochondrial structure. It is also involved in apoptosis.

There is increasing evidence that liver cell damage in chronic HBV infection is mediated by the induction of apoptosis. In apoptosis hyperphosphorylation of keratin filament occurs. Phosphorylated CK8/CK18 pair is the substrate for pro-caspase 3 and 9. These enzymes caspases become catalytically active and the effector caspases cleave CK-18. This keratin pairs in turn have been broken down and collapse the cytoplasmic and nuclear cytoskeleton. It leads to the condensation of chromatin, which is the hallmark of apoptosis. Caspase-cleaved CK18 fragment is released into the extracellular compartment. A monoclonal antibody, M30, specifically recognizes a fragment of CK18 cleaved at Asp396 (M30-antigen). An M30-based sandwich ELISA assay determines the circulating levels of M30-antigen and may serve as surrogate serum biomarker of hepatocyte apoptosis.¹²

METHODS AND MATERIALS:

Patients who were HBsAg +ve for more than 06 months with age- 18 to 65 years and HBV DNA value >2000 IU/ml were included in the study. Exclusion criteria were patient with HBsAg +ve for <6 months, co- infection

with HCV, HIV etc. patient with history of anti-viral treatment, alcohol consumption >30gm/day for male and >20gm/day for female, Non-alcoholic fatty liver disease, patient with decompensated cirrhosis of liver, patient with co-morbid condition (COPD, CKD, CCF etc.), patient who fails to give consent for biopsy. Total 40 patients were included after matching criteria.

Serum levels of M30-antigen were determined by commercially available immunoassays (M30-Apoptosense ELISA kit, Peviva AB, Bromma, Sweden) according to the manufacturer's instructions in the department of microbiology and immunology, BSMMU.

Per cutaneous transthoracic liver biopsy performed using a true-cut biopsy needle, G14, 15cm in length (sample length 1.5cm) with available resuscitation facilities. All biopsies were fixed with 10% formalin solution and stained with hematoxylin-eosin and Masson Trichrome stain. Experienced single pathologist, not aware about the clinical and biochemical parameters of any patient using the METAVIR scoring system evaluated biopsies. Patient were observed for 48 hours and then discharged from hospital.

Statistical Analysis

The demographic information, relevant history, examination findings and investigation reports of all the study subjects was recorded in previously prepared data collection sheet. After compilation, the data were presented in the form of tables, figures and graphs, as

necessary. Statistical analysis of the results as done by using computer based software, SPSS version.20 (SPSS Inc, Chicago, IL, USA).All values were presented as mean \pm standard deviations (SDs) for continuous data and as percentages for categorical data. Qualitative data were analyzed by Chi-square test and quantitative data were analyzed by student's t-test and anova test. Biochemical scoring was calculated using available formula. The AUROC, sensitivity, specificity and cut off values for biochemical indices were measured. A probability 'P' value of 0.05 or less was considered as significant. The association between serum biochemical indices with Fibrosis staging and the histological severity of CHB was evaluated by Spearman's correlation test.

RESULTS:

Table 1: Baseline characteristics of the study population (n=40)

Investigation	Range (min, max)	Mean \pm SD
Age (in years)	18, 50	26.5 \pm 7.7
Sex (male/female)-		31/9
Hb%	11.5, 16.2	13.7 \pm 1.3
TC	5, 12	7.4 \pm 1.7
Neutrophil (%)	41, 77	60.1 \pm 9.4
Lymphocyte (%)	18, 46	31.2 \pm 7.8

Monocyte (%)	1, 8	4.5 \pm 1.9
Eosinophil (%)	0, 15	4.2 \pm 3.0
Basophil (%)	0, 2	0.10 \pm 0.37
Platelet count ($\times 10^9/L$)	100, 400	234.3 \pm 70.1
*HBV DNA PCR (IU/ml)	2.2, 8.8	5.3 \pm 1.7
AST (UL)	19, 118	40.2 \pm 20.2
ALT (U/L)	22, 397	66.4 \pm 68.2
Prothrombin Time (sec)	10.5, 15.2	12.4 \pm 0.7
INR	0.88, 107	1.1 \pm 0.2
GGT	12, 110	30.2 \pm 17.9

All values are expressed as mean \pm SD or number (%).

Among the 40 patients, 16 patients were HBeAg positive and 24 patients were HBeAg negative.

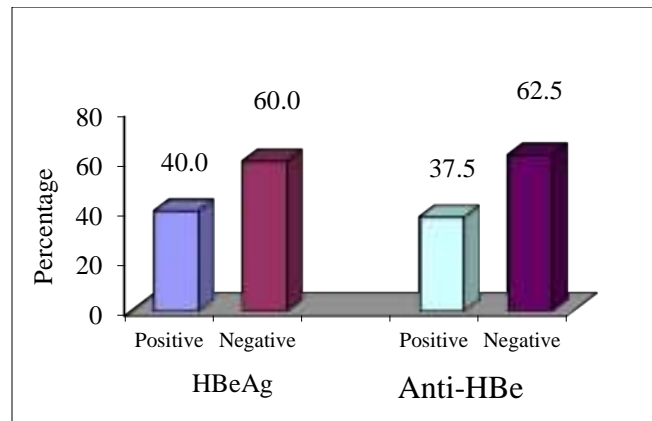


Fig. 1: Bar diagram shows HBeAg status of the study patients.

Table 2: Comparison of Demographic details and Laboratory findings between HBeAg positive and HBeAg negative CHB patients (n = 40).

HBeAg status	All patients (n = 40)	HBeAg Positive CHB (n = 16)	Negative CHB (n = 24)
*Age	26.5 \pm 7.7	24.9 \pm 9.2	27.6 \pm 6.7
**Sex (male/female)	31/9	13/3	17/7

Hb%	13.7±1.3	13.7±1.5	13.7±1.3
TC	7.4±1.7	7.1±1.7	7.7±1.7
Platelet count (*10 ⁹ /L)	234.3±70.1	218.5±45.1	244.8±81.9
*HBV DNA PCR (IU/ml)	5.3±1.7	7.0±0.9	4.2±1.0
AST (UL)	40.2±20.2	42.3±17.8	38.8±21.9
ALT (U/L)	66.4±68.2	62.1±41.5	69.3±82.0
Prothrombin Time (sec)	12.4±0.7	12.3±0.7	12.4±0.8
INR	1.1±0.2	1.1±0.6	1.2±0.2
GGT	30.2±17.9	32.6±23.6	28.6±13.2
Activity score			
A0	0 (0.0%)	0(0.0%)	0 (0.0%)
A1	7 (17.5%)	4 (25.0%)	3 (12.5%)
A2	21 (52.5%)	8 (50.0%)	13 (54.2%)
A3	12 (30.0%)	4 (25.0%)	8 (33.3%)
Fibrosis score			
F0	1 (2.5%)	0 (0.0%)	1 (4.2%)
F1	5 (12.5%)	2 (12.5%)	3 (12.5%)
F2	27 (67.5%)	10 (62.5%)	17 (70.8%)
F3	5 (12.5%)	3 (18.8%)	2 (8.3%)
F4	2 (5.0%)	1 (6.3%)	1 (4.2%)
Serum CK -18 fragment M30 level (Unit/L)	125.4±30.2	128.8±32.9	123.9±28.1

Table 03: Multiple comparison of variables in study patients according to Activity METAVIR score of hepatic necro inflammation (n=40)

	Activity score						P value
	A1		A2		A3		
	n	%	n	%	n	%	
Age (in years)	21.3	±4.6	27.7	±7.1	27.4	±9.3	0.145 ^{ns}
Sex (male/female)	5/2		15/6		10/2		0.728 ^{ns}
Prothombin time (sec)	12.3	±0.7	12.4	±0.7	12.3	±0.8	0.911 ^{ns}
Serum CK -18 fragment M30 level	140.6	±27.5	121.8	±25.1	124.4	±37.6	0.350 ^{ns}
AST	28.0	±8.4	38.5	±14.6	50.2	±28.5	0.055 ^{ns}
ALT	42.9	±14.0	53.1	±29.6	103.3	±112.2	0.072 ^{ns}
GGT	31.0	±35.3	29.4	±13.3	31.2	±11.7	0.956 ^{ns}
Platelet count	258.0	±61.5	237.9	±68.3	214.2	±77.6	0.407 ^{ns}
HBV DNA PCR	6.0	±2.1	5.1	±1.5	5.4	±1.6	0.457 ^{ns}

All values are expressed as mean±SD or number (%)

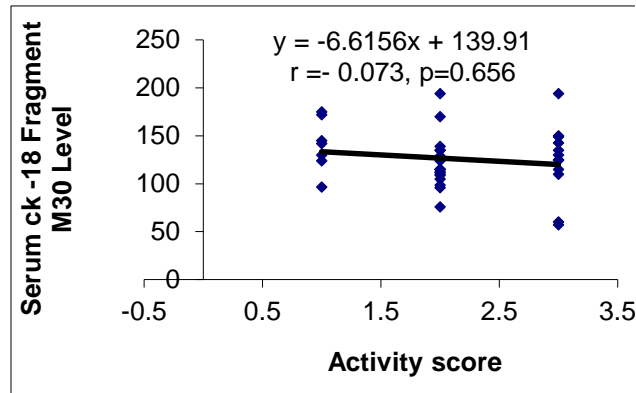


Fig. 2: Scatter diagram showing negative correlation ($r = -0.073$; $p = 0.357$) between activity score and serum CK -18 fragment M30 level.

Table 4: Correlation between factors and activity score of necro inflammation on spearman correlation test.

Factors	Correlation co-efficient value)	(r ^p -value)
Age	0.204	0.206 ⁿ _s
GGT	0.287	0.087 ⁿ _s
ALT	0.333	0.036 ^s
AST	0.360	0.016 ^s
Prothombin time	0.150	0.357 ⁿ _s
Serum CK -18 fragment M30 level	-0.073	0.656 ⁿ _s
HBV DNA PCR	-0.098	0.549 ⁿ _s
Platelet count	-0.289	0.070 ⁿ _s

DISCUSSION:

Increased CK-18 M30 level significantly correlates with the activity of hepatic necroinflammation and reflects as the non-invasive marker for significant inflammation (CB, Bae et al. 2013). So, measurement of serum CK-18 M 30 level can be useful for the predication of significant inflammation in CHB patients. In this study, most of the patients were young with 30 patients (75%) below the age of 30 years. Mean age of the patients found 26.5 ± 7.7 with the highest frequency at 21-30 years' age group (Table-1).

Among 40 CHB patients a male predominance was observed, male

31(77.5%) and female 9(22.5%). Among the 40 patients, 16 patients were HBeAg positive and 24 patients were HBeAg negative. Predominance of HBeAg negative patients was also similar in previous study done in Bangladeshi people.¹⁴In this study, comparison between HBeAg negative CHB patients and HBeAg positive patients reveals that HBeAg negative CHB patients are older (27.6 ± 6.7 versus 24.9 ± 9.2 years),AST and HBV DNA levels are lower (38.8 versus 42.3 U/L and 4.2 ± 1.0 Vs 7.0 ± 0.9 IU/ml respectively).They have significant fibrosis (82.2% Vs 75%) and significant inflammation (87.5% Vs 75%)(Table 2).These results are almost similar with another study conducted in Dept of

hepatology, BSMMU.¹⁵ In this study, CK-18 M30 level in both HBeAg negative CHB patients and HBeAg positive patients are almost similar (128.8±32.91 and 123.9±28.1) (Table 3). In this study, 17.5% patients have not significant necroinflammation (A0, A1) and 85.5% patients have significant necroinflammation (A2, A3). Serum CK-18 M30 level are not significantly increased in a stepwise fashion from A0 to A3. But AST and ALT levels are increased in a stepwise fashion from A0 to A3. Sherbiny WA et al., 2015 conducted a study in patients with chronic hepatitis C. Results showed that serum CK-18 M30 concentrations were significantly increased in a stepwise fashion from A0 to A3. Our study does not support this finding.

Correlation between factors and activity METAVIR score on spearman correlation test reveals, Serum CK-18 fragment M30 level was negatively correlated with activity score of hepatic necro inflammation ($r = -0.073$; $p = 0.357$). The correlation between ALT and AST with activity score were statistically significant ($p < 0.05$). Our study shows negative correlation with activity of necroinflammation.

The area under the receiver-operator characteristic (ROC) curves for prediction of serum CK -18 fragment M30 level reveals - area under curve(AUC) 0.307, which gave a cut off value of 100 U/L with 78.8% sensitivity and 14.3% specificity for prediction of significant necroinflammation (A2 and A3). Study conducted by Bae CB *et al.*, 2013 showed combined measurements of serum M30-antigen level (>344 U/L) and

AST (>78 IU/L) provided the most accurate identification of significant inflammation, showing 38.2% sensitivity, 96.1% specificity. Our study differs from that result.

CONCLUSION:

This study indicates there was no correlation between serum CK-18 fragment M30 level and hepatic necroinflammatory activity in patients with HBV related compensated chronic liver disease (CLD). As a result, CK 18 M30 cannot be used as an accurate non-invasive predictor of significant inflammatory activities in patients with CHB.

REFERENCE:

1. Global Hepatitis Report 2017. World Health Organization, 2017.
2. Schweitzer A, Horn J, Mikolajczyk RT, Krause G, Ott JJ. Estimations of worldwide prevalence of chronic hepatitis B virus infection: a systematic review of data published between 1965 and 2013. *Lancet*. 2015;386(10003):1546±55. [https://doi.org/10.1016/S0140-6736\(15\)61412-X](https://doi.org/10.1016/S0140-6736(15)61412-X) PMID: 26231459.
3. Te HS, Jensen DM. Epidemiology of hepatitis B and C viruses: a global overview. *Clinics in liver disease*. 2010; 14(1):1±21, vii. <https://doi.org/10.1016/j.cld.2009.11.009> PMID: 20123436
4. McMahon BJ. The natural history of chronic hepatitis B virus infection. *Hepatology*. 2009 May 1;49(S5):S45-55.

5. Hadziyannis SJ, Papatheodoridis GV. Hepatitis B e antigen-negative chronic hepatitis B: natural history and treatment. In Seminars in liver disease 2006 May (Vol. 26, No. 02, pp. 130-141). Copyright© 2006 by Thieme Medical Publishers, Inc., 333 Seventh Avenue, New York, NY 10001, USA.
6. Mahtab MA, Rahman S, Karim MF, Khan M, Foster G, Solaiman S, Afroz S. Epidemiology of hepatitis B virus in Bangladeshi general population. Hepatobiliary & pancreatic diseases international: HBPD INT. 2008 Dec;7(6):595-600.
7. Alam S, Ahmad N, Mustafa G, Alam K, Khan M. Characteristics of treatment naive chronic hepatitis B in Bangladesh: Younger populations are more affected; HBeAg-negatives are more advanced. Saudi journal of gastroenterology: official journal of the Saudi Gastroenterology Association. 2008 Jan;14(1):15.
8. Rahman S. Hepatitis B: From Blumberg to Bangladesh. Euroasian Journal of Hepato-Gastroenterology. 2012 Jan 25;1(2):42-3.
9. Fattovich G, Brollo L, Giustina G, Noventa F, Pontisso P, Alberti A, Realdi G, Ruol A. Natural history and prognostic factors for chronic hepatitis type B. Gut. 1991 Mar 1;32(3):294-8.
10. Ma J, Jiang Y, Gong G. Evaluation of seven noninvasive models in staging liver fibrosis in patients with chronic hepatitis B virus infection. European journal of gastroenterology & hepatology. 2013 Apr 1;25(4):428-34.
11. Chen B, Ye B, Zhang J, Ying L, Chen Y. RDW to platelet ratio: a novel noninvasive index for predicting hepatic fibrosis and cirrhosis in chronic hepatitis B. PloS one. 2013 Jul 17;8(7):e68780.
12. Erdogan S, Dogan HO, Sezer S, Uysal S, Ozhamam E, Kayacetin S, Koca Y. The diagnostic value of non-invasive tests for the evaluation of liver fibrosis in chronic hepatitis B patients. Scandinavian journal of clinical and laboratory investigation. 2013 Jun 1;73(4):300-8.
13. Mueller S, Seitz HK, Rausch V. Non-invasive diagnosis of alcoholic liver disease. World journal of gastroenterology: WJG. 2014 Oct 28;20(40):14626.
14. Mahtab MA, Rahman S, Khan M, Karim MF. Hepatitis E virus is a leading cause of acute-on-chronic liver disease: experience from a tertiary centre in Bangladesh. Hepatobiliary & pancreatic diseases international: HBPD INT. 2009 Feb;8(1):50-2.
15. Alam S, Ahmad N, Mustafa G, Shrestha A, Alam AK, Khan M. Evaluation of normal or minimally elevated alanine transaminase, age and DNA level in predicting liver histological changes in chronic hepatitis B. Liver international. 2011 Jul;31(6):824-30.